

# Spinal cord injury – there is not just one way of treating it

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## Abstract

In the last century, research in the field of spinal cord trauma has brought insightful knowledge which has led to a detailed understanding of mechanisms that are involved in injury- and recovery-related processes. The quest for a cure for the yet generally incurable condition as well as the exponential rise in gained information has brought about the development of numerous treatment approaches while at the same time the abundance of data has become quite unmanageable. Owing to an enormous amount of preclinical therapeutic approaches, this report highlights important trends rather than specific treatment strategies. We focus on current advances in the treatment of spinal cord injury and want to further draw attention to arising problems in spinal cord injury (SCI) research and discuss possible solutions.

## Regeneration in the central nervous system

For a very long time, the generally accepted hypothesis—initially proposed in the Edwin Smith papyrus in 2,550 BCE—had been that an injury of the spinal cord is an untreatable condition. In the 1920s, Ramon y Cajal postulated that central nervous system (CNS) axons have an intrinsic ability to regrow after injury but that the lack of trophic support and the barrier function of the lesion scar resulted in the observed lack of axonal regeneration after CNS trauma [1]. Despite almost a century of intensive investigation, the progress of therapeutic interventions to treat SCI remains very limited.

## Advances in spinal cord injury research

In the early 1980s, David and Aguayo reported that CNS axonal processes were able to regenerate for remarkable distances when provided the opportunity to grow through long peripheral nerve bridges circumventing a spinal cord lesion [2]. However, the respective nerve fibers did not leave the graft to re-enter the distal spinal cord, an observation that still holds for most therapeutic approaches in the field of SCI. Since then, diagnostic SCI imaging techniques have been immensely improved, and many important growth factors and receptors,

signaling cascades, cellular processes, and arising mechanisms of action have been investigated and characterized. Potential drug and cell treatment approaches have been developed based on these findings, and therapeutic interventions have been invented and successfully tested in preclinical studies. A few of these innovative therapies have already been subject to clinical trials, but the promising effects that were observed in preclinical animal studies could never be achieved to comparable degrees in human subjects [3].

Improved SCI outcome has been achieved by different repair strategies in animal models; the aim of neuroprotection strategies is to counteract secondary injury processes that lead to a progressive post-traumatic destruction of spinal cord tissue [4]. Examples of neuroprotective agents that have proven effective in animal models are the phosphodiesterase-4 inhibitor rolipram [5] and the Rho inhibitor cethrin [6]. Repair via regenerative sprouting can be achieved by (1) neutralization of inhibitory factors (e.g. the myelin protein Nogo-A [7]) or (2) administration of neurotrophic factors (e.g. neurotrophins [8]) or chemokines (e.g. CXCL12/SDF-1 [9,10]) to induce the intrinsic neuronal regeneration

program promoting axon growth and plasticity. On the other hand, those factors could influence axon regeneration and guidance through activation or inhibition of signaling pathways regulating the expression or activity of chemorepellent guidance molecules [11,12]. Axon regeneration can be further promoted by inhibition of (glial/fibrotic) scarring, either via degradation or suppression of inhibitory molecules [13,14] or by bridging the injury [15,16]. Although reactive astrocytes are often regarded to be detrimental to the functional outcome of SCI, these cells also mediate important protective functions [17]. Because astrocytes can play such dual roles in SCI response, the transplantation of specifically pre-differentiated astrocytes has also been proposed as a possible treatment for SCI and has been shown to promote functional recovery in animal models [18]. The death of oligodendrocytes, the myelinating cells of the CNS, is an acute result of SCI. In addition to axonal damage, demyelination leads to loss of axonal signal conduction and therefore to functional impairment in SCI. While mature oligodendrocytes are often lost to necrotic and apoptotic injury-related processes, a rapid proliferation and migration of oligodendrocyte precursor cells, especially at the lesion borders, can be observed [19]. For this reason, protected oligodendrocytic cells or their replacement or both are additional promising targets for therapeutic intervention after SCI. Promotion of remyelination is a key point in therapy development. Remyelination can be achieved by the induction of endogenous myelinating cells or by transplantation of myelin-producing cells [20,21]. Cell transplantation therapies have been developed to stimulate regenerative axon growth [22] or to replace lost cells and thereby repair the injured spinal cord [23], and gene therapeutic approaches with genetically modified cells or *in vivo* gene therapy can further support spinal cord repair [24]. Through transplantation of neural stem cells, lost glial and neuronal cells could be replaced, leading to remyelination [25] and axon regeneration [26] in animal models of SCI.

### Single-treatment approaches

Effective therapies are available for several SCI symptoms: brain stimulation can be used to treat neuropathic pain [27], and pharmacological treatments can act as neuroprotectants [28] or can reduce spasticity [29,30] or detrimental inflammation at the injury site [31]. Furthermore, therapeutic approaches to achieve improvements in quality of life by regaining breathing function [32], bladder/bowel function [33,34], or hand function [35] have proven quite successful in animal models of SCI. Existing treatment strategies as well as the majority of animal models generally target distinct symptoms of SCI, thereby neglecting the complexity of

a wide range of additional parameters. However, many of these approaches have delivered important insight into SCI pathology and have helped bring SCI therapies closer to feasibility. Examples are numerous approaches targeting the injury scar [13,14] to make it more permissive for regenerative axon growth [7,13,14,36,37], influencing inflammatory processes [38-40], peripheral nerve transplants [41-43], numerous cellular transplantation approaches [22,44-46], conditioning lesions [47], or gene therapy [48,49].

Several inhibitory molecules and some of their receptors have been identified and can be targeted by therapy. Some inhibitors of axonal growth are associated with white matter myelin (e.g. Nogo-A, myelin associated glycoprotein [MAG], and oligodendrocyte myelin glycoprotein [OMgp]) [50,51], whereas examples for scar-based inhibitory molecules which accumulate in the SCI lesion scar are members of the large class of chondroitin sulfate proteoglycans (CSPGs) [7,13]. Methods to block or disable the inhibitory function of these molecules have been developed. The myelin-associated inhibitor Nogo-A, for instance, can be neutralized by a specific antibody (IN-1)[7,52], and the glycosaminoglycan side chains of CSPGs can be degraded enzymatically by the bacterial enzyme chondroitinase ABC [7,13].

Strategies to improve locomotor function have demonstrated that small numbers of regenerating nerve fibers can suffice to achieve varying degrees of locomotor functional recovery [13,14,53]. At the same time, high numbers of regenerating nerve fibers—which can be achieved by cell transplantation—do not necessarily result in a considerably improved degree of functional recovery [26] when compared with “conventional” treatments [16,22]. On the contrary, regenerative axon growth, whether at a high or a low rate, could always result in adverse effects such as plastic changes leading to neuropathic pain [54,55]. Strategies to improve locomotor function have also shown that intrinsic spinal neuronal networks such as central pattern generators (CPGs) mediate certain aspects of locomotion even in the absence of sensory feedback [56,57]. CPGs are also important for functions like swallowing and breathing. Finally, (personalized) neuroprosthetics can aid in restoring locomotion even in the absence of axon regeneration and re-synaptogenesis. Therefore, such devices are very promising for future clinical applications [58-61].

### Questions of age and timing

Regeneration studies are generally performed in young adult animals, whereas the SCI epidemiology shows that there is an increased incidence at older ages [62] in

human patients. Older age seems to impair axonal plasticity rather than axon regeneration [63,64]. This finding could be of importance because it suggests that effective therapies to promote axon regeneration are still feasible for elderly patients, but it also suggests that different efficacies must be expected for treatments in young and aged patients. Additionally and very importantly, many therapies are being developed in acute injury models. However, a successful treatment for acute SCI may not be equally effective in sub-acute or even chronic SCI. The fibrous SCI scar develops in the first week after the insult [65]; therefore, many treatments target the lesion scar because it is a major physical and molecular barrier to regenerative axon growth. For a large number of treatments, the existing scar at the site of injury is an obstacle that needs to be overcome, especially in sub-acute and chronic SCI. There are different ways of approaching this matter: cellular or acellular matrices, filaments, or channels can be inserted, which function as bridging or guidance structures (or both) or deliver pharmaceutically active substances [15,16,66-69]. A very innovative treatment tool for this purpose is a recently described mechanical microconnector system [70], which actively reconnects severed spinal cord tissue stumps and can further deliver fluid pharmaceuticals into the lesion center cross-section, which is generally not well accessible for continuous *in situ* treatment. With this microconnector system, as with all experimental treatments, a major challenge will be the translation into large animal models up to the potential future clinical application where spinal defects will be of considerably larger size than in rodents. From a clinical point of view, a sub-acute therapy might be the most reasonable because patients with SCI might be reluctant to undergo further invasive interventions once they have come to terms with their situation. In this context, psychological care is absolutely vital to help the patient to accept his or her fate [71]. Regarding SCI in general and sub-acute and chronic SCI in particular, rehabilitative training requires a mention: it can increase neuronal plasticity and is one of the most successful current treatments [72] and, unlike most other SCI therapies, has made its way from the application in patients into experimental animal research (bedside-to-bench) and not *vice versa*. In the long term, locomotor training is of importance because, owing to muscle atrophy over time, axonal regeneration may not be as effective since respective functions must be re-learned. This makes rehabilitation an essential factor for chronic SCI research and treatment. Rehabilitation can be provided as either "passive exercise" or "active exercise". For passive exercise, volitional control is not mandatory. Repetitive training by passive exercise can modulate the caudal spinal circuitry to "normalize" specific spinal reflexes in

the absence of supraspinal control and thereby can reduce spasticity [73]. Active exercise can promote recovery by mediating plasticity at multiple levels of the neuraxis [73]. Active exercise requires varying degrees of supraspinal or spinal control or both. Unlike passive exercise, active exercise is, therefore, appropriate only after incomplete SCI. Whereas passive training can activate and increase joint motion, active exercise can improve motor recovery by further activating the muscles and multiple modes of afferent stimulation, possibly by changing the expression of inhibitory or supporting factors (or both) [74] or by altering electrophysiological properties in the lumbar enlargement [75].

### Combinatory treatments on the rise

Aside from obvious differences (e.g. spinal level, nature of the injury, and gross classification of the severity), no two cases of SCI are alike. Some single treatments can have multiple (additive or synergistic) effects (e.g. cell transplantation strategies [76,77], the administration of neurotrophic factors [78], or modulation of the SCI scar [13-15,79]). The multitude of research foci reflects the diversity of SCI. Therefore, the current trend is moving from single treatments to more holistic combinatory approaches [80]. As an example for a scar-modulating therapy, the bacterial enzyme chondroitinase ABC has been applied initially as a single treatment in rodents and currently is used in different small and large experimental animal studies [13,32,33,81,82] and also in numerous combinatory approaches [32,33,83-87]. At present, there are also numerous attempts to combine available pharmacological treatments with training in order to maximize the treatment effects. However, such combinatory strategies raise many new questions regarding, for example, the right timing (delay) to start the training, the intensity of the training sessions, or possible adverse effects [72]. Generally, a holistic combinatory SCI therapy should address as many aspects as possible, including neuroprotection (at early post-injury phases), the promotion of axonal growth, and modulating the lesion scar to make it more permissive for growing axons. Regular rehabilitative training should be a self-evident component of any SCI treatment. The major challenge in designing effective strategies will be the potential negative effects of the combination of generally beneficial single approaches. The development of such a comprehensive treatment requires intensive research and careful considerations to reduce the currently unmanageable number of potential treatment strategies.

### Managing the data overflow

The list of varying parameters and protocols in SCI research is already daunting (e.g. different injury models

in different species, strains, and sub-strains of experimental animals), and additionally taking into account the timing of treatment (acute, sub-acute, and chronic) and the recent trend toward combinatory treatments results in tremendous amounts of data, which are impossible to follow up by standard methods. This data overflow has recently led to an inevitable new focus in the field of SCI and in medicine in general: structuring, evaluating, and grading existent data in standardized ways. In this context, an international group of SCI researchers has joined to design new software and knowledge databases that allow the users to structure, manage, evaluate, and grade published data. Important examples are surveys, a grading system, and systematic reviews by Kwon and Tetzlaff and colleagues [88-91]; the Facilities of Research Excellence-Spinal Cord Injury (FORE-SCI) [92]; the Center for Neuronal Regeneration (CNR) [93]; and the reporting standard initiative Minimal Information About a Spinal Cord Injury Experiment (MIASCI) [94]. These initiatives have clearly demonstrated several problems in SCI research which result in a general lack of reproducibility. Problems in inter-lab reproducibility, presumably resulting from the above-mentioned varying protocols, have recently been disclosed. Influencing factors include small sample sizes (especially in many preclinical studies); lack of appropriate controls, of blinding, and of reliable outcome measures; and a general lack of transparency. On these grounds, the aforementioned initiatives address existing problems and try to bring potential solutions to the attention of the scientific community as well as the public. Furthermore, researchers are understandably reluctant to publish negative data. This leads to a serious lack of data availability. Overcoming this restraint could significantly accelerate the development of clinical therapies because it could prevent superfluous, time-consuming, and costly reproductions of negative data.

### Clinical trials – neuroprotection, functional repair, and regeneration

In addition to a network of scientific (see above) and clinical databases, e.g. the European Multicenter Study about Spinal Cord Injury (EM-SCI [95]), the successful translation of preclinical animal models requires the enforcement of strict criteria, which are established by regulatory agencies. Recently, various clinical trials based on innovative and novel experimental approaches have shown an efficacy in preclinical animal studies and safety in SCI patients but failed to show significant and reproducible efficacy [96]. One of the early clinical SCI trials was conducted by Proneuron Biotechnologies [97]. It comprised the transplantation of autologous (the patient's own) activated macrophages as a treatment for

acute SCI. The phase II study was suspended prematurely (supposedly not because of clinical or safety concerns). Some current/planned early-phase (phase I and II) clinical trials include conventional drug and molecule therapies such as the anti-Nogo A antibody therapy (antibody directed against an axon growth inhibitory protein in CNS myelin; desired effects: neuroprotection, axonal sprouting, and regeneration [98-100]) or cethrin treatment (cethrin acts as a C3 Rho inhibitor; desired effects: neuroprotection, axonal sprouting, and regeneration [101-103]). Cell transplantation approaches like the transplantation of autologous olfactory ensheathing glia (desired effects: axonal sprouting and regeneration [104,105]), of autologous bone marrow cells (desired effect: functional repair [20,106-109]), and of autologous Schwann cells (desired effect: functional repair [20,110,111]) are also very promising. However, in contrast to the transplantation of the latter cells, clinical approval of embryonic stem cells (ESCs) remains highly questionable because of the tumorigenic potential of ESC or ESC-derived precursor cells [112] in spite of potential beneficial treatment effects.

### Perspective

This short summary describing SCI symptoms, recent research approaches, and treatments up to current clinical applications is far from being complete. It highlights certain facets of a very complex disease pattern and outlines some attempts of aiding the patients' recovery. The hope remains that GLP (good laboratory practice) standardization of SCI studies, the availability and management of necessary data, and well-conducted clinical trials will eventually lead to the development of effective and, most likely, combinatorial treatments from which SCI patients can benefit.

### Abbreviations

CNS, central nervous system; CPG, central pattern generator; CSPG, chondroitin sulfate proteoglycan; ESC, embryonic stem cell; SCI, spinal cord injury.

### Disclosures

The authors declare that they have no disclosures.

### References

1. Ramon y Cajal, S: *Degeneration and regeneration of the nervous system*. New York: Oxford University Press; 1928.
2. David S, Aguayo AJ: **Axonal elongation into peripheral nervous system “bridges” after central nervous system injury in adult rats.** *Science* 1981, **214**:931-3.
3. Meurer WJ, Barsan WG: **Spinal Cord Injury Neuroprotection and the Promise of Flexible Adaptive Clinical Trials.** *World Neurosurg* 2013.



4. Becker D, McDonald JW: **Approaches to repairing the damaged spinal cord: overview.** *Handb Clin Neurol* 2012, **109**:445-61.
5. Nikulina E, Tidwell JL, Dai HN, Bregman BS, Filbin MT: **The phosphodiesterase inhibitor rolipram delivered after a spinal cord lesion promotes axonal regeneration and functional recovery.** *Proc Natl Acad Sci USA* 2004, **101**:8786-90.
6. McKerracher L, Higuchi H: **Targeting Rho to stimulate repair after spinal cord injury.** *J Neurotrauma* 2006, **23**:309-17.
7. Fawcett JW, Schwab ME, Montani L, Brazda N, Müller HW: **Defeating inhibition of regeneration by scar and myelin components.** *Handb Clin Neurol* 2012, **109**:503-22.
8. Hollis ER, Tuszyński MH: **Neurotrophins: potential therapeutic tools for the treatment of spinal cord injury.** *Neurotherapeutics* 2011, **8**:694-703.
9. Jaerve A, Bosse F, Müller HW: **SDF-1/CXCL12: its role in spinal cord injury.** *Int J Biochem Cell Biol* 2012, **44**:452-6.
10. Opatz J, Kury P, Schiwy N, Jarve A, Estrada V, Brazda N, Bosse F, Müller HW: **SDF-1 stimulates neurite growth on inhibitory CNS myelin.** *Mol Cell Neurosci* 2009, **40**:293-300.
11. Giger RJ, Hollis ER, Tuszyński MH: **Guidance molecules in axon regeneration.** *Cold Spring Harb Perspect Biol* 2010, **2**:a001867.
12. Niclou SP, Ehler EM, Verhaagen J: **Chemorepellent axon guidance molecules in spinal cord injury.** *J Neurotrauma* 2006, **23**:409-21.
13. Bradbury EJ, Moon LD, Popat RJ, King VR, Bennet GS, Patel PN, Fawcett JW, McMahon SB: **Chondroitinase ABC promotes functional recovery after spinal cord injury.** *Nature* 2002, **416**:636-40.

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14. Klapka N, Hermann S, Straten G, Masanneck C, Duis S, Hamers FP, Frank P T, Müller D, Zuschratter W, Müller HW: **Suppression of fibrous scarring in spinal cord injury of rat promotes long-distance regeneration of corticospinal tract axons, rescue of primary motoneurons in somatosensory cortex and significant functional recovery.** *Eur J Neurosci* 2005, **22**:3047-58.
15. Estrada V, Brazda N, Schmitz C, Heller S, Blazyca H, Martini R, Müller HW: **Long-lasting significant functional improvement in chronic severe spinal cord injury following scar resection and polyethylene glycol implantation.** *Neurobiol Dis* 2014, **67C**:165-79.
16. Sakiyama-Elbert S, Johnson PJ, Hodgetts SI, Plant GW, Harvey AR: **Scaffolds to promote spinal cord regeneration.** *Handb Clin Neurol* 2012, **109**:575-94.
17. Faulkner JR, Herrmann JE, Woo MJ, Tansey KE, Doan NB, Sofroniew MV: **Reactive astrocytes protect tissue and preserve function after spinal cord injury.** *J Neurosci* 2004, **24**:2143-2155.

**F1000Prime  
RECOMMENDED**

18. Davies SJ, Shih CH, Noble M, Mayer-Proschel M, Davies JE, Proschel C: **Transplantation of specific human astrocytes promotes functional recovery after spinal cord injury.** *PLoS One* 2011, **6**:e17328.

**F1000Prime  
RECOMMENDED**

19. Almad A, Sahinkaya FR, McTigue DM: **Oligodendrocyte fate after spinal cord injury.** *Neurotherapeutics* 2011, **8**:262-273.
20. Guest J, Santamaria AJ, Benavides FD: **Clinical translation of autologous Schwann cell transplantation for the treatment of spinal cord injury.** *Curr Opin Organ Transplant* 2013, **18**:682-89.

**F1000Prime  
RECOMMENDED**

21. Nakamura M, Okano H: **Cell transplantation therapies for spinal cord injury focusing on induced pluripotent stem cells.** *Cell Res* 2013, **23**:70-80.
22. Schira J, Gasis M, Estrada V, Hendricks M, Schmitz C, Trapp T, Kruse F, Köller G, Wernet P, Hartung H, Müller HW: **Significant clinical, neuropathological and behavioural recovery from acute spinal cord trauma by transplantation of a well-defined**

**somatic stem cell from human umbilical cord blood.** *Brain* 2012, **135**:431-46.

23. Cao Q, Whittemore SR: **Cell transplantation: stem cells and precursor cells.** *Handb Clin Neurol* 2012, **109**:551-61.
24. Franz S, Weidner N, Blesch A: **Gene therapy approaches to enhancing plasticity and regeneration after spinal cord injury.** *Exp Neurol* 2012, **235**:62-69.
25. Yasuda A, Tsuji O, Shibata S, Nori S, Takano M, Kobayashi Y, Takahashi Y, Fujiyoshi K, Hara CM, Miyawaki A, Okano H, Toyama Y, Nakamura M, Okano H: **Significance of remyelination by neural stem/progenitor cells transplanted into the injured spinal cord.** *Stem Cells* 2011, **29**:1983-94.

**F1000Prime  
RECOMMENDED**

26. Lu P, Wang Y, Graham L, McHale K, Gao M, Wu D, Brock J, Blesch A, Rosenzweig ES, Havton LA, Zheng B, Conner JM, Marsala M, Tuszyński MH: **Long-distance growth and connectivity of neural stem cells after severe spinal cord injury.** *Cell* 2012, **150**:1264-73.

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RECOMMENDED**

27. Nardone R, Holler Y, Leis S, Holler P, Thon N, Thomschewski A, Golaszewski S, Brigo F, Trinka E: **Invasive and non-invasive brain stimulation for treatment of neuropathic pain in patients with spinal cord injury: A review.** *J Spinal Cord Med* 2014, **37**:19-31.
28. Garrido-Mesa N, Zarzuelo A, Galvez J: **Minocycline: far beyond an antibiotic.** *Br J Pharmacol* 2013, **169**:337-52.
29. Dietz V, Sinkjaer T: **Spasticity.** *Handb Clin Neurol* 2012, **109**:197-211.
30. McIntyre A, Mays R, Mehta S, Janzen S, Townsend A, Hsieh J, Wolfe D, Teasell R: **Examining the effectiveness of intrathecal baclofen on spasticity in individuals with chronic spinal cord injury: A systematic review.** *J Spinal Cord Med* 2014, **37**:11-8.

**F1000Prime  
RECOMMENDED**

31. Juknis N, Cooper JM, Volshteyn O: **The changing landscape of spinal cord injury.** *Handb Clin Neurol* 2012, **109**:149-66.
32. Alilain WJ, Horn KP, Hu H, Dick TE, Silver J: **Functional regeneration of respiratory pathways after spinal cord injury.** *Nature* 2011, **475**:196-200.

**F1000Prime  
RECOMMENDED**

33. Lee YS, Lin CY, Jiang HH, Depaul M, Lin VW, Silver J: **Nerve regeneration restores supraspinal control of bladder function after complete spinal cord injury.** *J Neurosci* 2013, **33**:10591-606.

**F1000Prime  
RECOMMENDED**

34. Pan Y, Liu B, Li R, Zhang Z, Lu L: **Bowel Dysfunction in Spinal Cord Injury: Current Perspectives.** *Cell Biochem Biophys* 2014, **69**:385-8.
35. Nishimura Y, Perlmutter SI, Fetz EE: **Restoration of upper limb movement via artificial corticospinal and musculospinal connections in a monkey with spinal cord injury.** *Front Neural Circuits* 2013, **7**:57.
36. Cregg JM, Depaul MA, Filous AR, Lang BT, Tran A, Silver J: **Functional regeneration beyond the glial scar.** *Exp Neurol* 2014, **253C**:197-207.
37. Silver J, Miller J: **Regeneration beyond the glial scar.** *Nat Rev Neurosci* 2004, **5**:146-156.

**F1000Prime  
RECOMMENDED**

38. Benowitz LI, Popovich PG: **Inflammation and axon regeneration.** *Curr Opin Neurol* 2011, **24**:577-83.
39. Kigerl KA, Gensel JC, Ankeny DP, Alexander JK, Donnelly DJ, Popovich PG: **Identification of two distinct macrophage subsets with divergent effects causing either neurotoxicity**

**or regeneration in the injured mouse spinal cord.** *J Neurosci* 2009, **29**:13435-44.



40. Popovich PG, Longbrake EE: **Can the immune system be harnessed to repair the CNS?** *Nat Rev Neurosci* 2008, **9**:481-93.
41. Cote MP, Amin AA, Tom VJ, Houle JD: **Peripheral nerve grafts support regeneration after spinal cord injury.** *Neurotherapeutics* 2011, **8**:294-303.
42. Fraidakis MJ, Spenger C, Olson L: **Partial recovery after treatment of chronic paraplegia in rat.** *Exp Neurol* 2004, **188**:33-42.
43. Hill CE, Brodak DM, Bartlett BM: **Dissociated predegenerated peripheral nerve transplants for spinal cord injury repair: a comprehensive assessment of their effects on regeneration and functional recovery compared to Schwann cell transplants.** *J Neurotrauma* 2012, **29**:2226-43.
44. Li J, Lepski G: **Cell transplantation for spinal cord injury: a systematic review.** *Biomed Res Int* 2013, **2013**:786475.
45. Franz S, Weidner N, Blesch A: **Gene therapy approaches to enhancing plasticity and regeneration after spinal cord injury.** *Exp Neurol* 2012, **235**:62-9.
46. Hou T, Wu Y, Wang L, Liu Y, Zeng L, Li M, Long Z, Chen H, Li Y, Wang Z: **Cellular prostheses fabricated with motor neurons seeded in self-assembling peptide promotes partial functional recovery following spinal cord injury in rats.** *Tissue Eng Part A* 2012, **18**:974-85.
47. Blesch A, Lu P, Tsukada S, Alto LT, Roet K, Coppola G, Geschwind D, Tuszyński MH: **Conditioning lesions before or after spinal cord injury recruit broad genetic mechanisms that sustain axonal regeneration: superiority to camp-mediated effects.** *Exp Neurol* 2012, **235**:162-73.
48. Blesch A, Fischer I, Tuszyński MH: **Gene therapy, neurotrophic factors and spinal cord regeneration.** *Handb Clin Neurol* 2012, **109**:563-74.
49. Verhaagen J, Van Kesteren RE, Bossers KA, Macgillivray HD, Mason MR, Smit AB: **Molecular target discovery for neural repair in the functional genomics era.** *Handb Clin Neurol* 2012, **109**:595-616.
50. Filbin MT: **Myelin-associated inhibitors of axonal regeneration in the adult mammalian CNS.** *Nat Rev Neurosci* 2003, **4**:703-13.



51. Lee JK, Zheng B: **Role of myelin-associated inhibitors in axonal repair after spinal cord injury.** *Exp Neurol* 2012, **235**:33-42.
52. Caroni P, Schwab ME: **Antibody against myelin-associated inhibitor of neurite growth neutralizes nonpermissive substrate properties of CNS white matter.** *Neuron* 1988, **1**:85-96.
53. Fawcett J: **Repair of spinal cord injuries: where are we, where are we going?** *Spinal Cord* 2002, **40**:615-23.
54. Ferguson AR, Huie JR, Crown ED, Baumbauer KM, Hook MA, Garraway SM, Lee KH, Hoy KC, Grau JW: **Maladaptive spinal plasticity opposes spinal learning and recovery in spinal cord injury.** *Front Physiol* 2012, **3**:399.
55. Nardone R, Holler Y, Brigo F, Seidl M, Christova M, Bergmann J, Golaszewski S, Trinka E: **Functional brain reorganization after spinal cord injury: systematic review of animal and human studies.** *Brain Res* 2013, **1504**:58-73.
56. Musienko P, Heutschi J, Friedli L, van den BR, Courtine G: **Multisystem neurorehabilitative strategies to restore motor functions following severe spinal cord injury.** *Exp Neurol* 2012, **235**:100-9.
57. Edgerton VR, Roy RR: **Robotic training and spinal cord plasticity.** *Brain Res Bull* 2009, **78**:4-12.
58. Borton D, Micera S, Millan JR, Courtine G: **Personalized neuromodulation.** *Sci Transl Med* 2013, **5**:210rv2.



59. Borton D, Bonizzato M, Beauparlant J, DiGiovanna J, Moraud EM, Wenger N, Musienko P, Minev IR, Lacour SP, Millán, José del R, Micera S, Courtine G: **Corticospinal neuroprostheses to restore locomotion after spinal cord injury.** *Neurosci Res* 2014, **78**:21-9.



60. Courtine G, Micera S, DiGiovanna J, Millan JR: **Brain-machine interface: closer to therapeutic reality?** *Lancet* 2013, **381**:515-17.



61. van den BR, Heutschi J, Barraud Q, DiGiovanna J, Bartholdi K, Huerlimann M, Friedli L, Vollenweider I, Moraud EM, Duis S, Dominici N, Micera S, Musienko P, Courtine G: **Restoring voluntary control of locomotion after paralyzing spinal cord injury.** *Science* 2012, **336**:1182-85.



62. Johns Hopkins University School of Medicine. [http://www.hopkinsmedicine.org]
63. Wirz M, Dietz V: **Concepts of aging with paralysis: implications for recovery and treatment.** *Handb Clin Neurol* 2012, **109**:77-84.
64. Jaerve A, Schiwy N, Schmitz C, Müller HW: **Differential effect of aging on axon sprouting and regenerative growth in spinal cord injury.** *Exp Neurol* 2011, **231**:284-94.
65. Klapka N, Müller HW: **Collagen matrix in spinal cord injury.** *J Neurotrauma* 2006, **23**:422-35.
66. Hejcl A, Jendelova P, Sykova E: **Experimental reconstruction of the injured spinal cord.** *Adv Tech Stand Neurosurg* 2011, **37**:65-95.
67. Joosten EA: **Biodegradable biomaterials and bridging the injured spinal cord: the corticospinal tract as a proof of principle.** *Cell Tissue Res* 2012, **349**:375-95.
68. Kubinova S, Sykova E: **Biomaterials combined with cell therapy for treatment of spinal cord injury.** *Regen Med* 2012, **7**:207-24.
69. Tyler JY, Xu XM, Cheng JX: **Nanomedicine for treating spinal cord injury.** *Nanoscale* 2013, **5**:8821-36.
70. Brazda N, Voss C, Estrada V, Lodin H, Weinrich N, Seide K, Müller J, Müller HW: **A mechanical microconnector system for restoration of tissue continuity and long-term drug application into the injured spinal cord.** *Biomaterials* 2013, **34**:10056-64.

71. Grundy D, Swain A (eds.): *ABC of spinal cord injury* BMJ Books; 2002.
72. Fouad K, Tetzlaff W: **Rehabilitative training and plasticity following spinal cord injury.** *Exp Neurol* 2012, **235**:91-9.
73. Lynskey JV, Belanger A, Jung R: **Activity-dependent plasticity in spinal cord injury.** *J Rehabil Res Dev* 2008, **45**:229-40.
74. Houle JD, Cote MP: **Axon regeneration and exercise-dependent plasticity after spinal cord injury.** *Ann N Y Acad Sci* 2013, **1279**:154-63.
75. Beaumont E, Kaloustian S, Rousseau G, Cormery B: **Training improves the electrophysiological properties of lumbar neurons and locomotion after thoracic spinal cord injury in rats.** *Neurosci Res* 2008, **62**:147-54.
76. Garbossa D, Boido M, Fontanella M, Fronda C, Ducati A, Vercelli A: **Recent therapeutic strategies for spinal cord injury treatment: possible role of stem cells.** *Neurosurg Rev* 2012, **35**:293-311.
77. Ruff CA, Wilcox JT, Fehlings MG: **Cell-based transplantation strategies to promote plasticity following spinal cord injury.** *Exp Neurol* 2012, **235**:78-90.
78. Weishaupt N, Blesch A, Fouad K: **BDNF: the career of a multifaceted neurotrophin in spinal cord injury.** *Exp Neurol* 2012, **238**:254-64.
79. Barritt AW, Davies M, Marchand F, Hartley R, Grist J, Yip P, McMahon SB, Bradbury EJ: **Chondroitinase ABC promotes sprouting of intact and injured spinal systems after spinal cord injury.** *J Neurosci* 2006, **26**:10856-67.
80. Oudega M, Bradbury EJ, Ramer MS: **Combination therapies.** *Handb Clin Neurol* 2012, **109**:617-36.

81. Bowes C, Massey JM, Burish M, Cerkevich CM, Kaas JH: **Chondroitinase ABC promotes selective reactivation of somatosensory cortex in squirrel monkeys after a cervical dorsal column lesion.** *Proc Natl Acad Sci USA* 2012, **109**:2595-600.
- F1000Prime  
RECOMMENDED**
82. Garcia-Alias G, Lin R, Akrimi SF, Story D, Bradbury EJ, Fawcett JW: **Therapeutic time window for the application of chondroitinase ABC after spinal cord injury.** *Exp Neurol* 2008, **210**:331-8.
83. Wang D, Ichiyama RM, Zhao R, Andrews MR, Fawcett JW: **Chondroitinase combined with rehabilitation promotes recovery of forelimb function in rats with chronic spinal cord injury.** *J Neurosci* 2011, **31**:9332-44.
- F1000Prime  
RECOMMENDED**
84. Kanno H, Pressman Y, Moody A, Berg R, Muir EM, Rogers JH, Ozawa H, Ito I, Pearse DD, Bunge MB: **Combination of engineered Schwann cell grafts to secrete neurotrophin and chondroitinase promotes axonal regeneration and locomotion after spinal cord injury.** *J Neurosci* 2014, **34**:1838-55.
85. Soleman S, Filippov MA, Dityatev A, Fawcett JW: **Targeting the neural extracellular matrix in neurological disorders.** *Neuroscience* 2013, **253**:194-213.
86. Zhao RR, Andrews MR, Wang D, Warren P, Gullo M, Schnell L, Schwab ME, Fawcett JW: **Combination treatment with anti-Nogo-A and chondroitinase ABC is more effective than single treatments at enhancing functional recovery after spinal cord injury.** *Eur J Neurosci* 2013, **38**:2946-61.
87. Zhao RR, Fawcett JW: **Combination treatment with chondroitinase ABC in spinal cord injury-breaking the barrier.** *Neurosci Bull* 2013, **29**:477-83.
88. Kwon BK, Okon EB, Plunet W, Baptiste D, Fouad K, Hillyer J, Weaver LC, Fehlings MG, Tetzlaff W: **A systematic review of directly applied biologic therapies for acute spinal cord injury.** *J Neurotrauma* 2011, **28**:1589-610.
- F1000Prime  
RECOMMENDED**
89. Kwon BK, Okon E, Hillyer J, Mann C, Baptiste D, Weaver LC, Fehlings MG, Tetzlaff W: **A systematic review of non-invasive pharmacologic neuroprotective treatments for acute spinal cord injury.** *J Neurotrauma* 2011, **28**:1545-88.
- F1000Prime  
RECOMMENDED**
90. Kwon BK, Okon EB, Tsai E, Beattie MS, Bresnahan JC, Magnuson DK, Reier PJ, McTigue DM, Popovich PG, Blight AR, Oudega M, Guest JD, Weaver LC, Fehlings MG, Tetzlaff W: **A grading system to evaluate objectively the strength of pre-clinical data of acute neuroprotective therapies for clinical translation in spinal cord injury.** *J Neurotrauma* 2011, **28**:1525-43.
- F1000Prime  
RECOMMENDED**
91. Tetzlaff W, Okon EB, Karimi-Abdolrezaee S, Hill CE, Sparling JS, Plemel JR, Plunet WT, Tsai EC, Baptiste D, Smithson LJ, Kawaja MD, Fehlings MG, Kwon BK: **A systematic review of cellular transplantation therapies for spinal cord injury.** *J Neurotrauma* 2011, **28**:1611-82.
- F1000Prime  
RECOMMENDED**
92. Steward O, Popovich PG, Dietrich WD, Kleitman N: **Replication and reproducibility in spinal cord injury research.** *Exp Neurol* 2012, **233**:597-605.
- F1000Prime  
RECOMMENDED**
93. The Center for Neuronal Regeneration. [http://www.cnr.de]
94. Lemmon VP, Ferguson AR, Popovich PG, Xu XM, Snow DM, Igarashi M, Beattie CE, Bixby JL: **Minimum Information about a Spinal Cord Injury Experiment (MIASCI) - A Proposed Reporting Standard for Spinal Cord Injury Experiments.** *J Neurotrauma* 2014, **31**:1354-61.
95. van Hedel HJ, Wirz M, Dietz V: **Standardized assessment of walking capacity after spinal cord injury: the European network approach.** *Neuro Res* 2008, **30**:61-73.
96. Steeves J, Blight A: **Spinal cord injury clinical trials translational process, review of past and proposed acute trials with reference to recommended trial guidelines.** *Handb Clin Neurol* 2012, **109**:386-98.
97. **Proneuron Biotechnologies.** [http://www.proneuron.com]
98. Buchli AD, Schwab ME: **Inhibition of Nogo: a key strategy to increase regeneration, plasticity and functional recovery of the lesioned central nervous system.** *Ann Med* 2005, **37**:556-67.
99. Freund P, Schmidlin E, Wannier T, Bloch J, Mir A, Schwab ME, Rouiller EM: **Anti-Nogo-A antibody treatment promotes recovery of manual dexterity after unilateral cervical lesion in adult primates-re-examination and extension of behavioral data.** *Eur J Neurosci* 2009, **29**:983-96.
100. Zörner B, Schwab ME: **Anti-Nogo on the go: from animal models to a clinical trial.** *Ann N Y Acad Sci* 2010, **1198 Suppl 1**:E22-E34.
101. McKerracher L, Guertin P: **Rho as a target to promote repair: translation to clinical studies with cethrin.** *Curr Pharm Des* 2013, **19**:4400-410.
102. McKerracher L, Anderson KD: **Analysis of recruitment and outcomes in the phase I/IIa Cethrin clinical trial for acute spinal cord injury.** *J Neurotrauma* 2013, **30**:1795-804.
103. Lord-Fontaine S, Yang F, Diep Q, Dergham P, Munzer S, Tremblay P, McKerracher L: **Local Inhibition of Rho Signaling by Cell-Permeable Recombinant Protein BA-210 Prevents Secondary Damage and Promotes Functional Recovery following Acute Spinal Cord Injury.** *J Neurotrauma* 2008, **25**:1309-322.
104. Tabakow P, Jarmundowicz W, Czapiga B, Fortuna W, Miedzybrodzki R, Czyz M, Huber J, Szarek D, Okurowski S, Szewczyk P, Gorski A, Raisman G: **Transplantation of autologous olfactory ensheathing cells in complete human spinal cord injury.** *Cell Transplant* 2013, **22**:1591-612.
105. Mackay-Sim A, Feron F, Cochrane J, Bassingthwaite L, Bayliss C, Davies W, Fronek P, Gray C, Kerr G, Licina P, Nowitzke A, Perry C, Silburn, P A S, Urquhart S, Geraghty T: **Autologous olfactory ensheathing cell transplantation in human paraplegia: a 3-year clinical trial.** *Brain* 2008, **131**:2376-86.
106. Saito F, Nakatani T, Iwase M, Maeda Y, Hirakawa A, Murao Y, Suzuki Y, Onodera R, Fukushima M, Ide C: **Spinal cord injury treatment with intrathecal autologous bone marrow stromal cell transplantation: the first clinical trial case report.** *J Trauma* 2008, **64**:53-9.
107. Saito F, Nakatani T, Iwase M, Maeda Y, Murao Y, Suzuki Y, Fukushima M, Ide C: **Administration of cultured autologous bone marrow stromal cells into cerebrospinal fluid in spinal injury patients: a pilot study.** *Restor Neurol Neurosci* 2012, **30**:127-36.
108. Vaquero J, Zurita M: **Cell transplantation in paraplegic patients: the importance of properly assessing the spinal cord morphology.** *Clin Transplant* 2013, **27**:968-71.
109. Vaquero J, Zurita M: **Bone marrow stromal cells for spinal cord repair: a challenge for contemporary neurobiology.** *Histol Histopathol* 2009, **24**:107-16.
110. Guest JD, Hiester ED, Bunge RP: **Demyelination and Schwann cell responses adjacent to injury epicenter cavities following chronic human spinal cord injury.** *Exp Neural* 2005, **192**:384-93.
111. Bunge MB: **Novel combination strategies to repair the injured mammalian spinal cord.** *J Spinal Cord Med* 2008, **31**:262-9.
112. Sahni V, Kessler JA: **Stem cell therapies for spinal cord injury.** *Nat Rev Neurol* 2010, **6**:363-72.